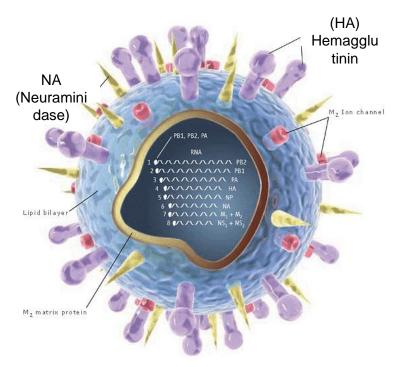


Unit 4
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INFLUENZA A VIRUS

- Negative sense, segmented RNA virus
- Orthomyxoviridae
- Eight genes, 11 proteins (three alternate reading frames)
- Two non-structural proteins (NS1 and PB1-F2)
- Surface proteins HA and NA determine serotype



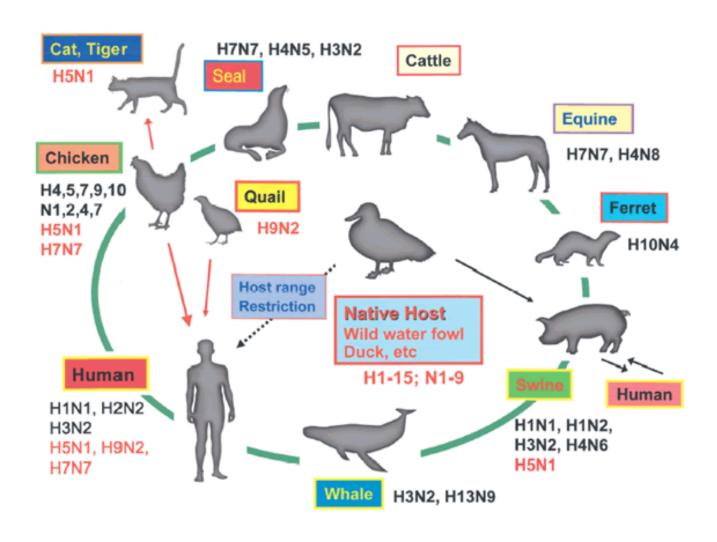
Modified from: Kaiser. Science 2006, 312:380-382.

Influenza A HA and NA Subtypes

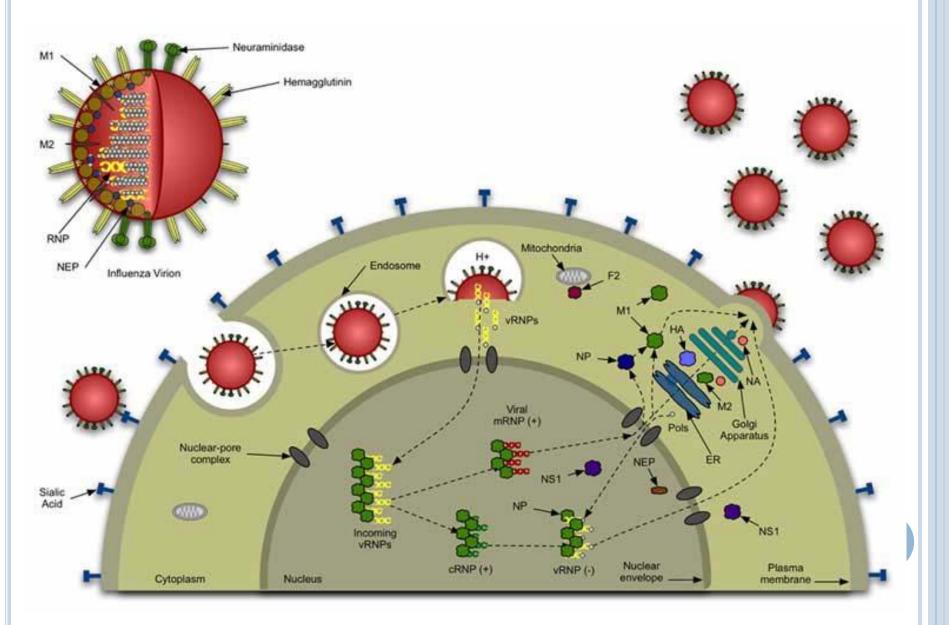
H1		200	
H2		200	
Н3		50	Other Animals
H4		50	Other Animals
H5		50	Other Animals
H6		**	
H7			Other Animals
H8			
H9		50	
H10		**	
H11			
H12			
H13			
H14			
H15			
H16			

N1		
N2		
N3		
N4		
N5		
N6		
N7		Other Animals
N8		Other Animals
N9		

DIVERSE HOST TROPISM ALLOWS RESTRICTION AND RECOMBINATION

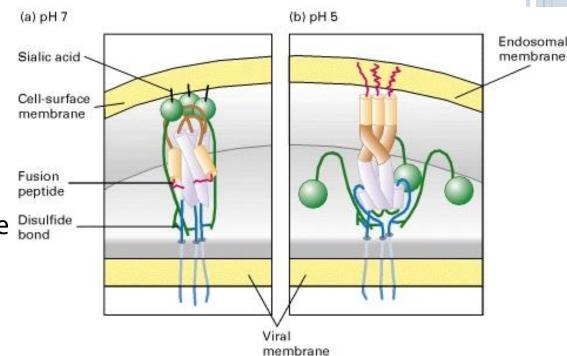


INFLUENZA LIFE CYCLE



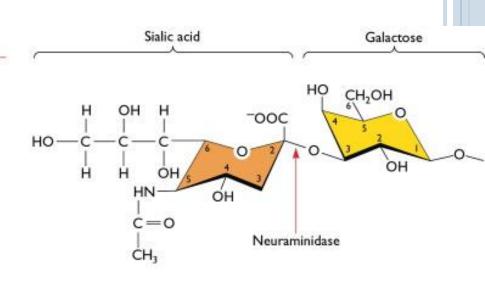
HA IS REQUIRED FOR CELL ENTRY

- HA binding to sialic acid on the surface of cells mediates initial attachment
- Virus is endocytosed, where the endosome is acidified
- This triggers a conformational change in the virus, resulting in membrane fusion
- For HA to be active, it needs to be cleaved by a protease into two pieces—this protease is generally restricted to the respiratory epithelium



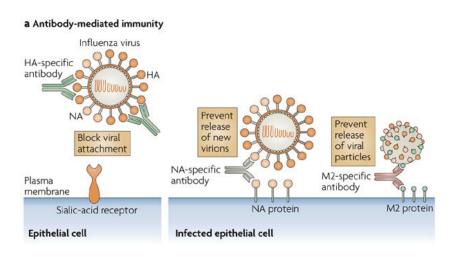
NEURAMINIDASE ACTS TO CLEAVE THE SIALIC ACID RECEPTORS FROM THE CELL SURFACE

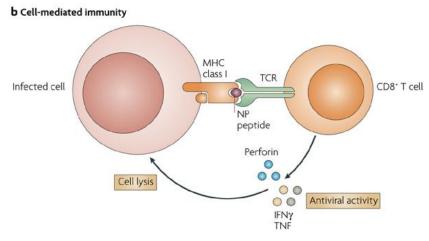
 IAV must balance the binding and entry activity of HA with the sialic acid cleavage activity of NA so that virus efficiently enters and buds from the cell surface—thus HA and NA are often "matched" for activity



IMMUNE MECHANISMS OF PROTECTION

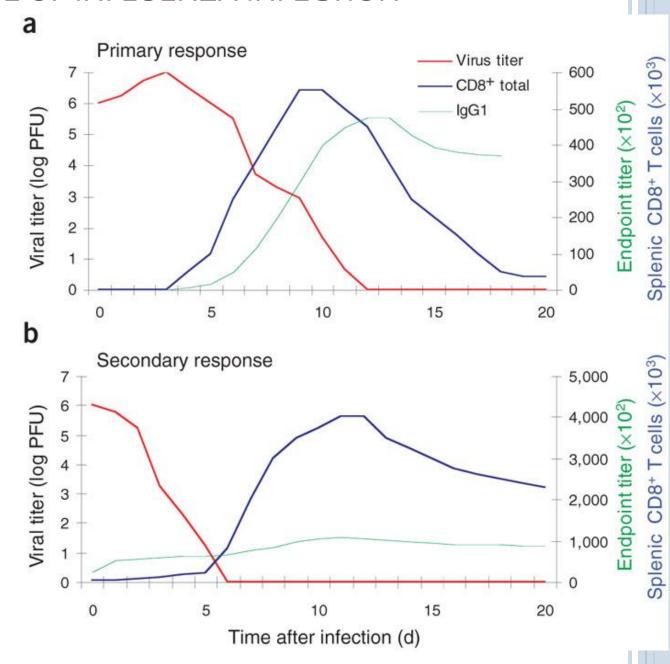
- Antibody mediated immunity exerts the most pressure on the virus, leading to seasonal antigenic drift and pandemic strains of antigenic shift
- Internal proteins are relatively conserved allowing heterologous cellular protection
- Mutation of dominant CD8 epitopes over time suggests that CTLs provide immunological pressure



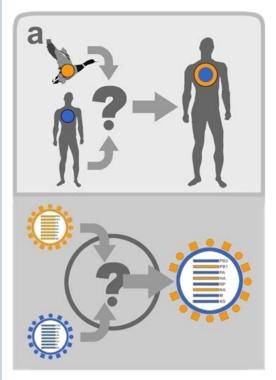


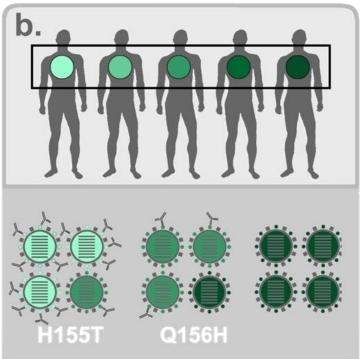
IMMUNE COURSE OF INFLUENZA INFECTION

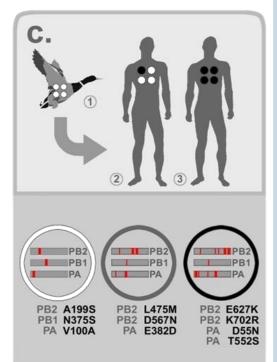
- Influenza is initially controlled by antibody and CD8+ T cells
- Secondary infection with heterologous virus is cleared with CD8+ T cell activity much more rapidly
- Homologous infection can be prevented by antibody (sterilizing immunity)



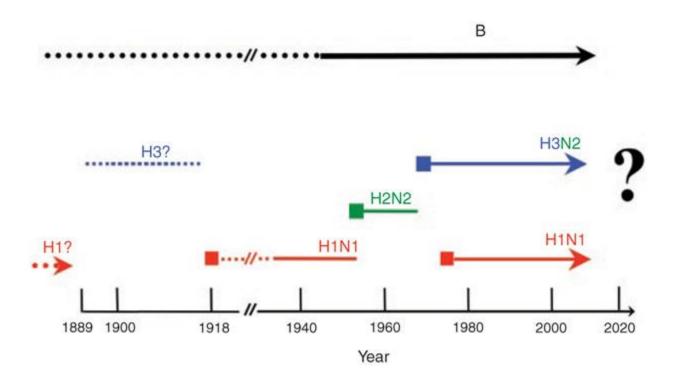
INFLUENZA EVOLUTION





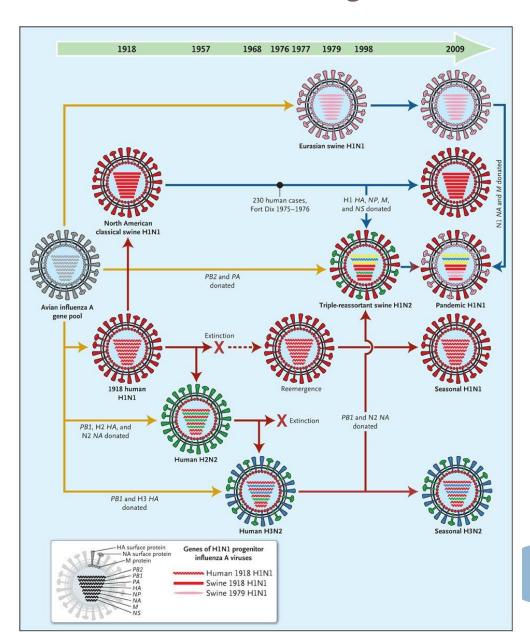


HUMAN INFLUENZA PANDEMICS



EVOLUTION OF HUMAN INFLUENZA FROM 1918

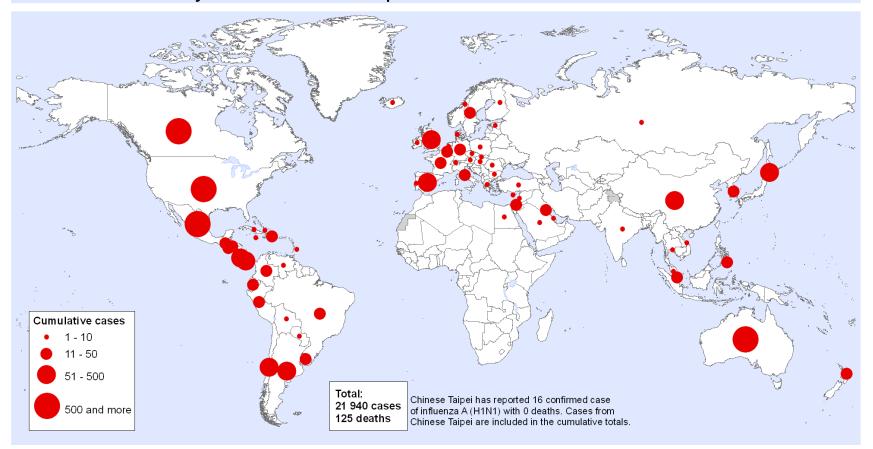
- All current human influenza is majorityderived from the 1918 pandemic
- Distinct reservoirs have allowed evolution to occur with varying pressures, providing diverse sources for new gene introductions into the human pool



SWINE-ORIGIN H₁N₁ INCIDENCE

New Influenza A (H1N1), Number of laboratory confirmed cases as reported to WHO

Status as of 05 June 2009 06:00 GMT



The boundaries and names shown and the designations used on this map do not imply the expression of any opinion whatsoever on the part of the World Health Organization concerning the legal status of any country, territory, city or area or of its authorities, or concerning the delimitation of its frontiers or boundaries. Dotted lines on maps represent approximate border lines for which there may not yet be full agreement.

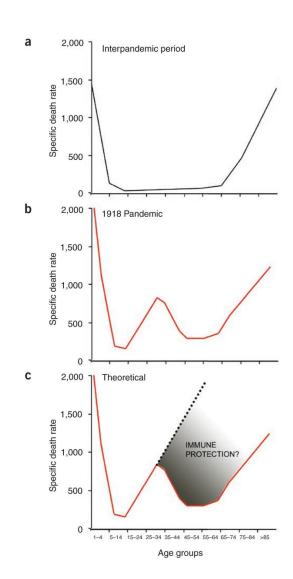
Data Source: World Health Organization
Map Production: Public Health Information
and Geographic Information Systems (GIS)
World Health Organization



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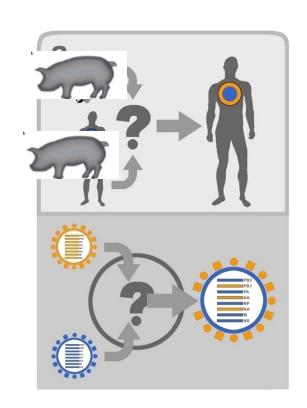
1918 (AND POSSIBLY SWORH1N1) MORTALITY CURVES SUGGEST PREVIOUS EXPOSURE

- The "U" shaped curve of regular influenza infection demonstrates the highest mortality among children (naïve) and the elderly (immunocomprimised)
- The 1918 pandemic had a "W" shaped curve, with a spike in deaths among young adults immunopathology or prior protection for ~40 year olds?



PREDICTIONS OF THE 2009/H1N1 PANDEMIC

- The 2009 H1N1 pandemic emerged as a particularly novel threat: an antigenic shift event between two swine viruses, without the "human" virus component expected to be required
- The initial rapid spread bred fears of an equally high incidence of severe morbidity and mortality (~90,000 deaths in the US, ~1.8 million hospitalizations)



PRE-EXISITNG CROSS-REACTIVE IMMUNITY TO 2009/H1N1

ORIGINAL ARTICLE

Cross-Reactive Antibody Responses to the 2009 Pandemic H1N1 Influenza Virus

Kathy Hancock, Ph.D., Vic Veguilla, M.P.H., Xiuhua Lu, M.D., Weimin Zhong, Ph.D., Eboneé N. Butler, M.P.H., Hong Sun, M.D., Feng Liu, M.D., Ph.D., Libo Dong, M.D., Ph.D., Joshua R. DeVos, M.P.H., Paul M. Gargiullo, Ph.D., T. Lynnette Brammer, M.P.H., Nancy J. Cox, Ph.D., Terrence M. Tumpey, Ph.D., and Jacqueline M. Katz, Ph.D.

Type of Vaccine, Influenza Season, and Influenza Virus Used in Assay	Age y Group	No. of Subjects	Increase in Antibody Titer by a Factor of ≥4	Geometric Mean Titer†		Microneutralization Titer of ≥40 for Children or ≥160 for Adults;	
				Before Vaccination (95% CI)	After Vaccination (95% CI)	Before Vaccination	After Vaccination
			%			9	6
Children							
Trivalent inactivated influenza va	accine						
2005–2007	6 mo to 9 yr	33					
Seasonal H1N1			67	26 (16–40)	267 (171–418)	45	94
Pandemic H1N1			0	5 (5–6)	6 (5–6)	0	0
2007–2008	5 yr to 9 yr	13					
Seasonal H1N1			85	42 (22–80)	575 (303–1093)	54	100
Pandemic H1N1			0	10 (7–15)	12 (8–17)	8	15
2008-2009	6 mo to 23 mo	9					
Seasonal H1N1			100	5 (4–7)	285 (202–402)	0	100
Pandemic H1N1§			0	5	5	0	0
Frivalent inactivated influenza va with adjuvant	accine						
2008-2009	6 mo to 59 mo	45¶					
Seasonal H1N1			96	12 (8–18)	193 (134–280)	24	100
Pandemic H1N1			2	6 (5–7)	8 (7–9)	0	4

TABLE CONTINUED

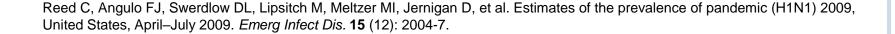
				(,)	(,-2)		
Adults							
Trivalent inactivated influenza v	accine						
2007–2008	18 yr to 64 yr	148					
Seasonal H1N1			75	48 (40–58)	598 (497–720)	29	93
Pandemic H1N1			22	25 (21–31)	54 (44–65)	7	25
2008-2009	18 yr to 40 yr	83					
Seasonal H1N1			78	29 (22–38)	546 (418–713)	20	88
Pandemic H1N1			12	11 (9–14)	21 (16–26)	6	7
Older adults							
Trivalent inactivated influenza vaccine							
2007-2008	≥60 yr	63					
Seasonal H1N1			54	31 (22–42)	143 (105–194)	14	54
Pandemic H1N1			5	92 (71–121)	97 (74–127)	33	43
2008-2009	≥60 yr						
Seasonal H1N1		49**	18	22 (17–28)	51 (39–66)	6	14
Pandemic H1N1		50**	0	47 (36–61)	51 (39–65)	8	8

EARLY PANDEMIC H1N1: APRIL – JULY 2009

Table 2. Estimates of pandemic (H1N1) 2009–related cases and rates of illness and hospitalization by age distribution of confirmed case-patients, United States, April–July 2009

	Estimate	ed no. case-patients	Estimated rate/100,000	
Parameter	Median	90% range	Median	90% range
Total no. case-patients by age group, y†	3,052,768	1,831,115-5,720,928	997	598-1,868
0–4	397,033	238,149-744,045	1,870	1,122-3,505
5–24	1,820,284	1,091,845-3,411,237	2,196	1,317–4,115
25–49	612,862	367,608–1,148,511	577	346-1,081
50–64	180,297	108,146-337,879	319	192-599
<u>></u> 65	42,292	25,368-79,256	107	64-201
No. hospitalized case-patients by age group, y	13,764	9,278-21,305	4.5	3.0-7.0
0–4	2,768	1,866-4,285	13.0	8.8-20.2
5–24	4,991	3,364–7,725	6.0	4.1-9.3
25-49	3.440	2.319-5.324	3.2	2.2-5.0
50–64	1,912	1,289-2,959	3.4	2.3-5.2
<u>≥</u> 65	654	441-1,012	1.7	1.1-2.6
Multiplier				
Hospitalized	2.7	1.7–4.5	-	_
Nonhospitalized	79	47 –148	_	_
Through May 12	33	23–49	_	_
After May 12	84	50-163	_	_

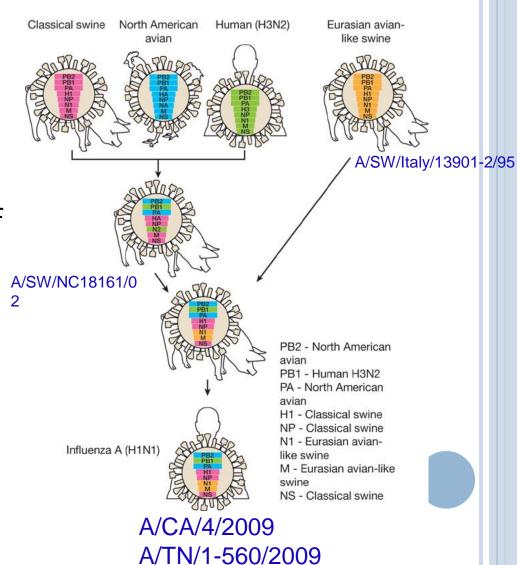
^{*}United States Population Estimates, 2009.



[†]Age distributions from line list and aggregate reports of laboratory-confirmed cases and hospitalizations to the Centers for Disease Control and Prevention through July 23, 3009.

2009 PANDEMIC H1N1

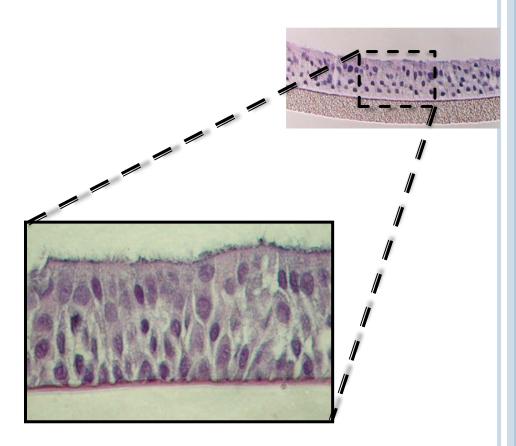
- 2009/H1N1 resulted from the recombination of two viruses (American and Eurasian Swine)
- The American Swine virus was itself a recombinant of three viruses that established itself in 1998
- These viruses are genetically distant from the human seasonal H1N1 (reference strain A/Brisbane/59/07)



H1N1 SWINE FLU STUDIES: RESPONSE IN HUMAN CELLS

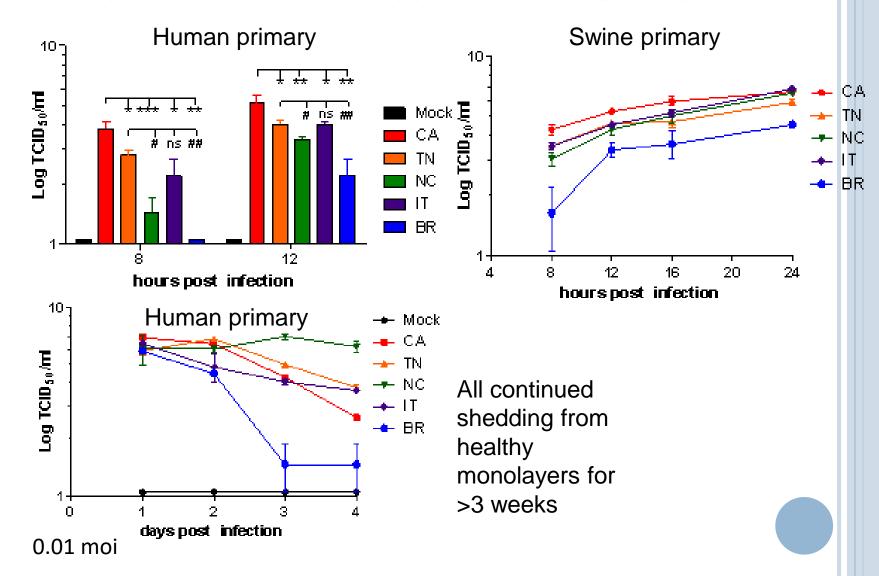
Measures:

- Infectivity and growth of virus (TCID₅₀, immunofluorescence)
- Secretion of inflammatory mediators from apical and basolateral surfaces (multiplexed immunoassay)
- Transcriptional response over the first 24 hours (Exon arrays, fluidigm analysis)
- Confirm results by "swapped viruses" made by reverse genetics

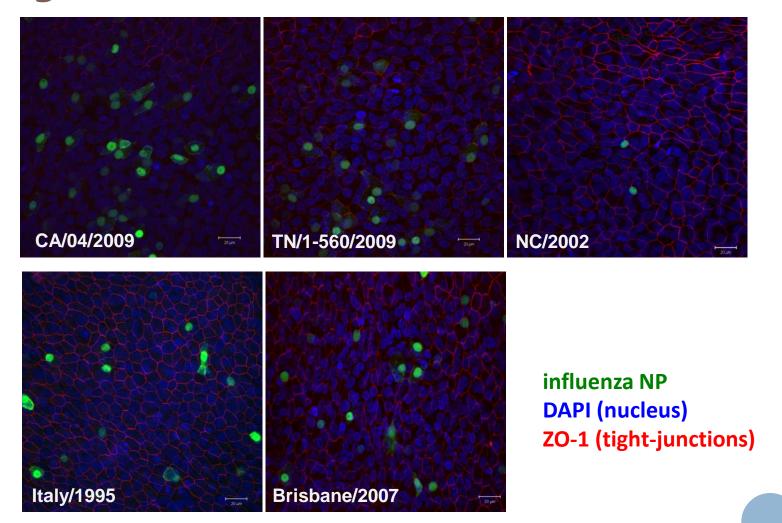


EpiAirway™, MatTek

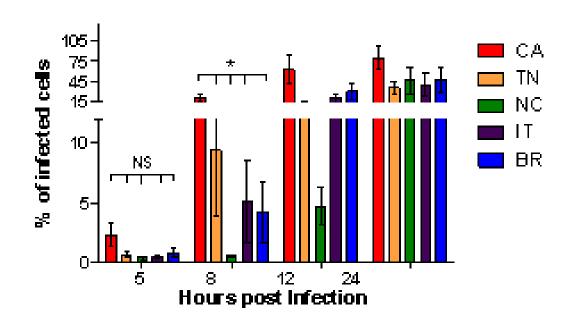
VIRAL GROWTH KINETICS IN HAE CELLS



Influenza NP detection in 3D HAE cultures viral growth kinetics in HAE cells



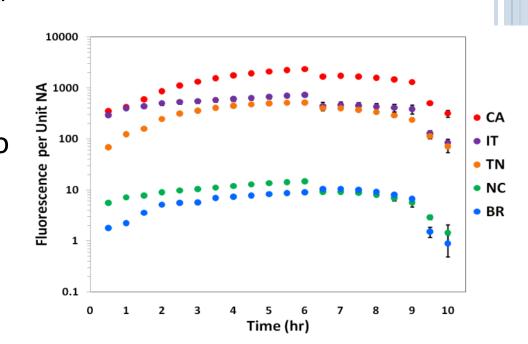
MORE RAPID COLONIZATION OF CULTURE BY PANDEMIC AND ESW VIRUS



By 12 hours, pandemic strains and Italy have infected ~50%-75% of the culture

HIGHER NA ACTIVITY IN PANDEMIC AND ESW

- NA activity measured as ability to convert sialic acid containing substrate
- Results normalized to functional viral titer, so NA activity/infectious virion
- Higher NA activity may relate to ability of virus to spread efficiently



GROWTH SUMMARY

- The pandemic virus acquired a rapid growth phenotype in human cells similar to the Esw virus
- This phenotype associates with both the NA and M of Esw virus
- The Esw virus transmits more efficiently in ferrets
- Titer and infected cell number can be de-coupled across infections/individuals

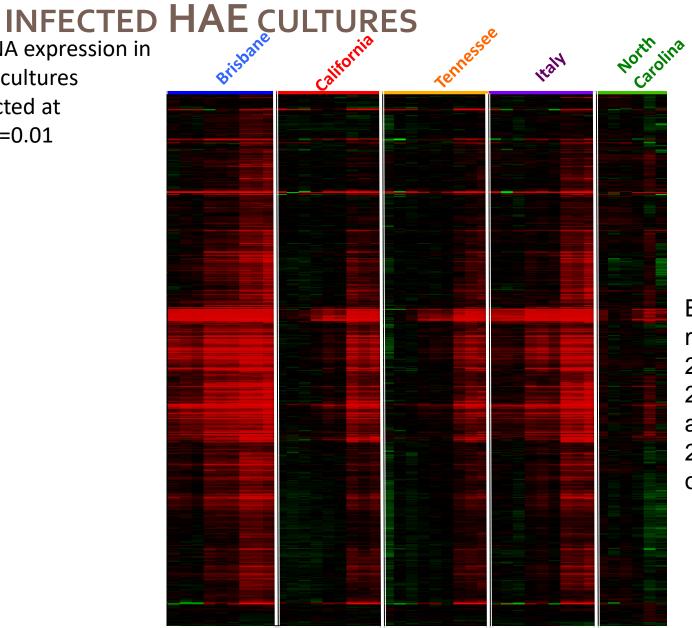
ODE MODEL OF INFLUENZA INFECTION—ANDREAS HANDEL, UGA

$$\begin{array}{lll} \frac{dU}{dt} & = & \lambda D - \frac{b}{1+s_1X}UV & \text{uninfected cells} \\ \frac{dE}{dt} & = & \frac{b}{1+s_1X}UV - \frac{g}{1+s_3X}E & \text{latent infected cells} \\ \frac{dI}{dt} & = & \frac{g}{1+s_3X}E - dI & \text{productively infected cells} \\ \frac{dD}{dt} & = & dI - \lambda D & \text{dead cells} \\ \frac{dV}{dt} & = & \frac{p}{1+s_2X}I - cV - \gamma \frac{b}{1+s_1X}VU & \text{free virus} \end{array}$$

Why wasn't the Esw virus a pandemic?

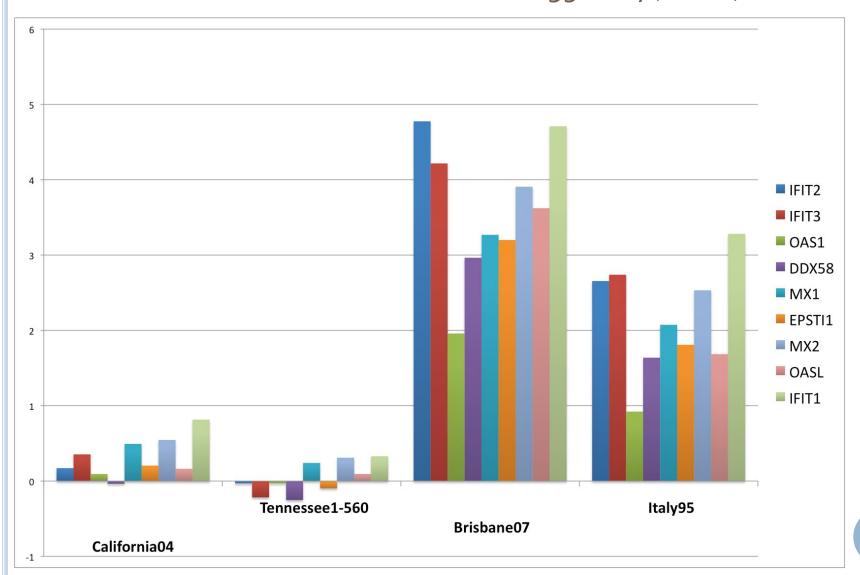
TRANSCRIPTOME ANALYSIS OF PANDEMIC VIRUS

mRNA expression in hAE cultures infected at MOI=0.01

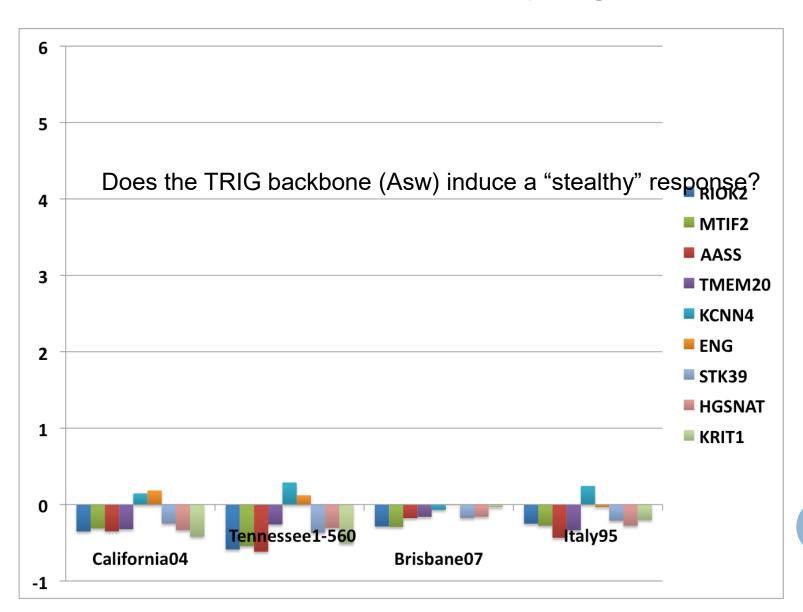


BIC applied to kmeans clustering: 2 clusters 271 upregulated in all 24 downregulated or differential

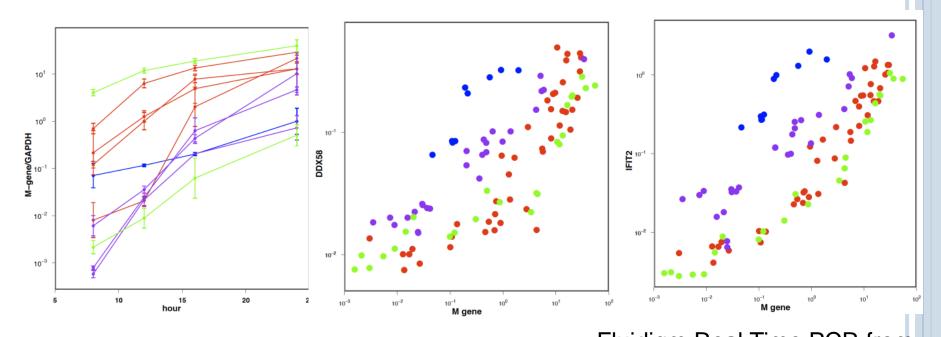
Top 9 Most significant differentially expressed genes 12 Hours post-infection with A/Brisbane/59/2007(H1N1)



TOP 9 MOST SIGNIFICANT DIFFERENTIALLY EXPRESSED GENES AT 12 HOURS POST-INFECTION WITH A/CALIFORNIA/04/2009(H1N1)



HOST RESPONSE AS A FUNCTION OF VIRUS



Fluidigm Real Time PCR from primary human cell infections (2 donors)

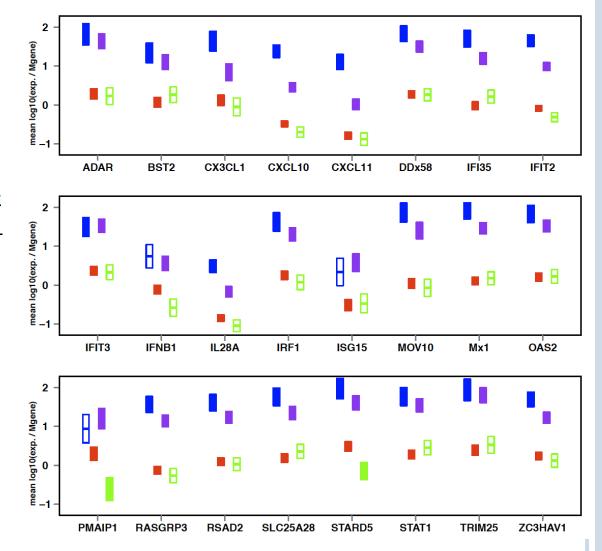
Brisbane California Italy

North Carolina

HOST RESPONSE AS A FUNCTION OF VIRUS II

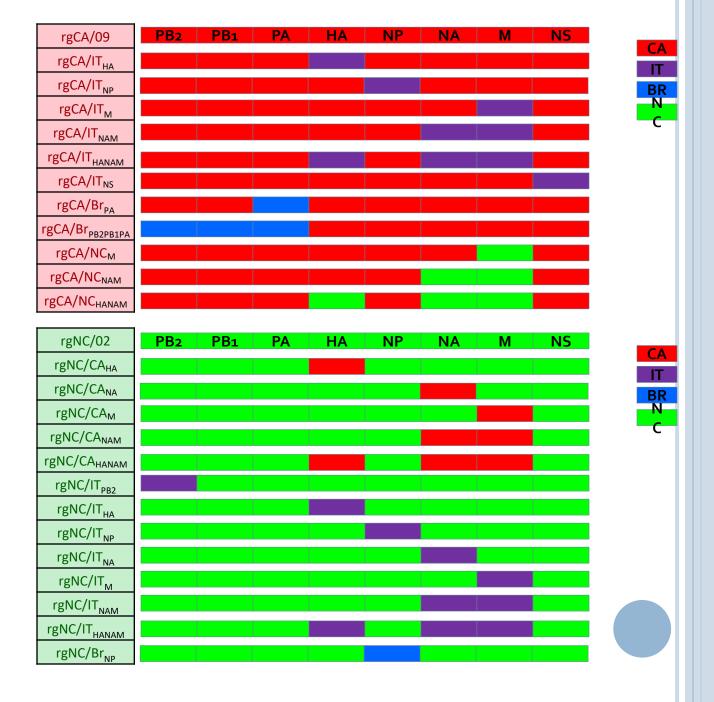
Brisbane California
North
Carolina

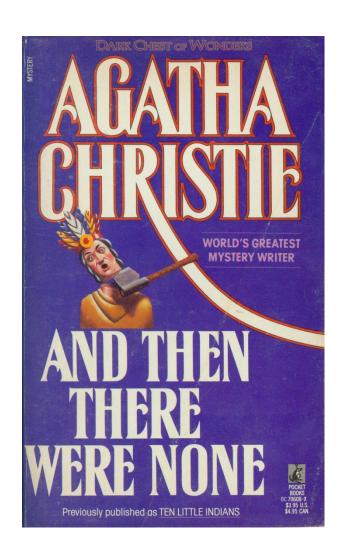
 $\frac{expression - expression_{mock}}{\max{(expression)}}$ Mgene

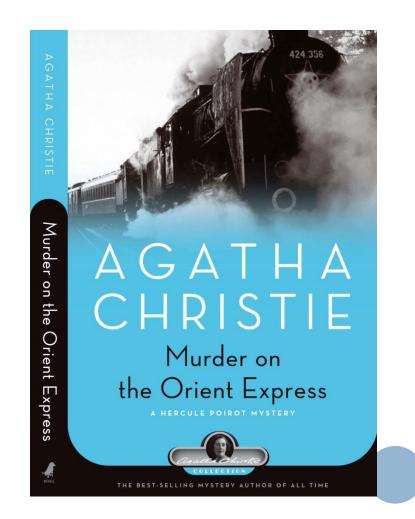


SWAPS

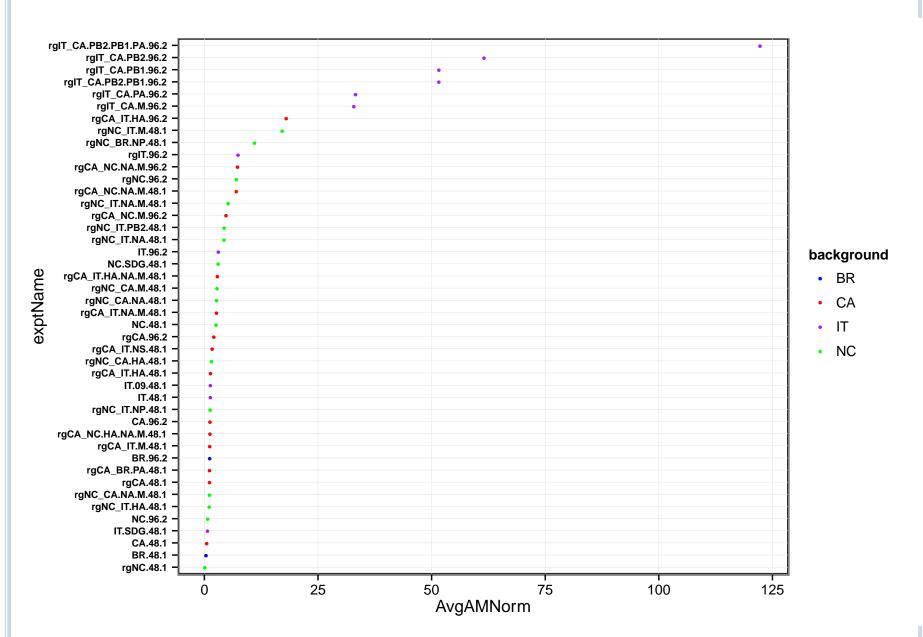
What's the mechanistic basis of the stealthy (or noisy) phenotype?



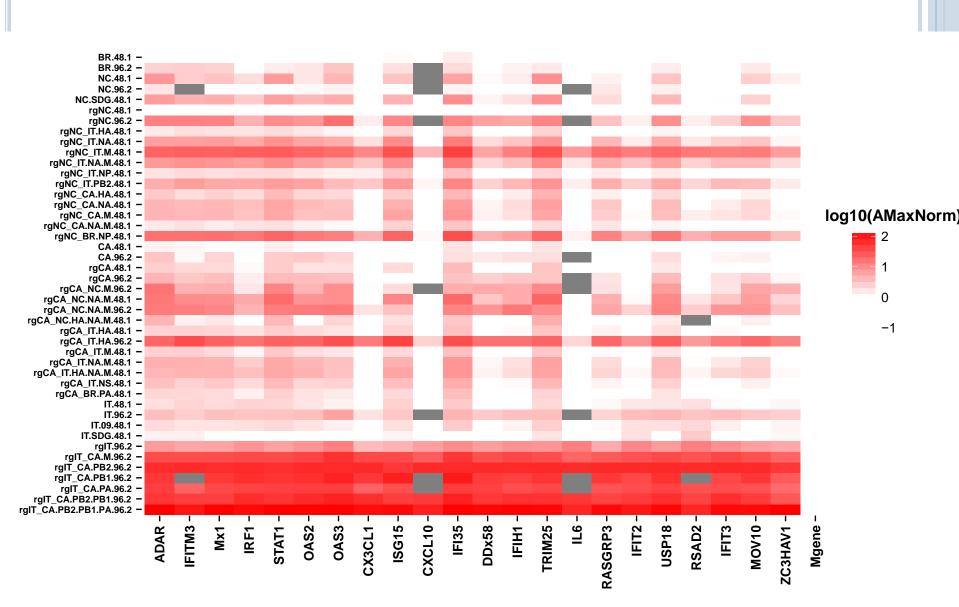




Average amplitude across all genes normalized to M-gene



Amplitude ("A") normalized to M-gene



THE PANDEMIC STRAIN IS EFFICIENT AND STEALTHY

Rapid + stealthy growth = Pandemic

Morbidity and Mortality Weekly Report

Limited Human-to-Human Transmission of Novel Influenza A (H3N2) Virus — Iowa, November 2011

- The set of genes induced by diverse viruses is largely equivalent in the first 24 hours— "the flu program"
- The pandemic strategy is distinct from the well-adapted human seasonal virus
- Kinetic differences in the first ~18 hours of infection are critical to the quality and quantity of the later response
- The stealthy phenotype ismediated by contributions of the P-gene complex, with potential roles for NP and NS

ODE MODEL OF INFLUENZA INFECTION

$$\begin{array}{lll} \frac{dU}{dt} &=& \lambda D - \frac{b}{1+s_1X}UV & \text{uninfected cells} \\ \frac{dE}{dt} &=& \frac{b}{1+s_1X}UV - \frac{g}{1+s_3X}E & \text{latent infected cells} \\ \frac{dI}{dt} &=& \frac{g}{1+s_3X}E - dI & \text{productively infected cells} \\ \frac{dD}{dt} &=& dI - \lambda D & \text{dead cells} \\ \frac{dV}{dt} &=& \frac{p}{1+s_2X}I - cV - \gamma \frac{b}{1+s_1X}VU & \text{free virus} \\ \frac{dX}{dt} &=& wI - \delta X & \text{innate immune response (IFN)} \end{array}$$

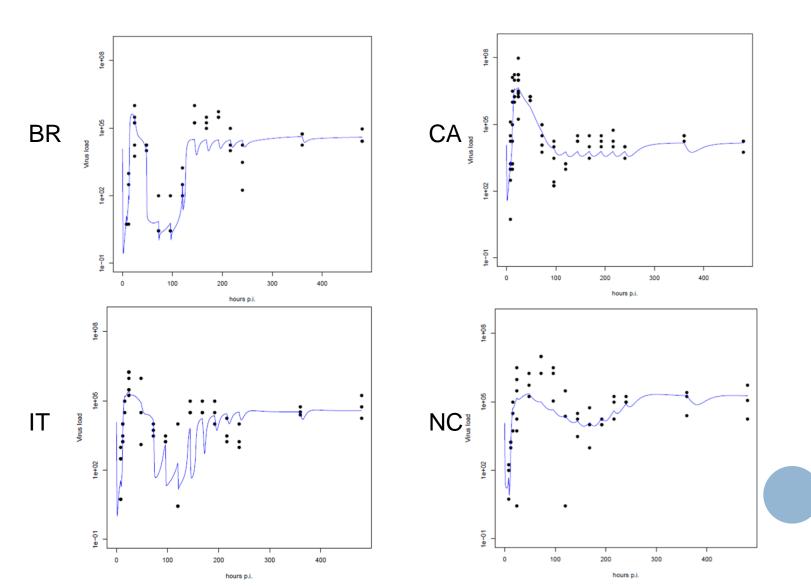
AICC VALUES OF 8 DIFFERENT MODELS

- 1. No IR and no cell-regrowth
- 2. No IR, with cell-regrowth
- With IR reducing virus production, no cellregrowth
- With IR reducing infection rate, no cellregrowth

- regrowth
- 6. With IR reducing virus production, with cell-regrowth
- With IR reducing infection rate, with cellregrowth
- 8. With IR prolonging latency, with cellregrowth

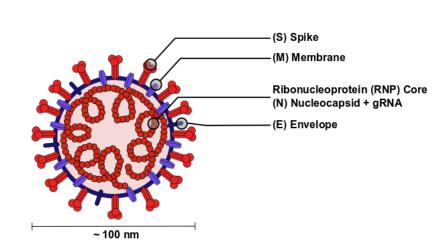
5. With Refelon	ging later R. no cell-	CA	\mathbf{IT}	NC
1	54.5	54.7	33.1	28.2
2	48.8	-22.6	0.8	28.5
3	52.8	24.8	17.0	30.3
4	59.9	33.2	38.3	33.6
5	53.2	32.1	24.6	31.7
6	-11.6	-17.6	-11.1	33.2
7	54.5	-17.7	6.1	29.3
8	56.1	-17.3	6.2	34.3

FITS FOR MODEL 6—IR REDUCES VIRUS PRODUCTION AND CELLS REGROW



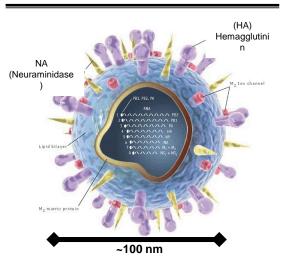
SARS-COV-2 VS. INFLUENZA VIRUS

The Coronavirus Virion



(+) ssRNA genome ~28-32 Kb 29 proteins

The Influenza Virus Virion



(-) segmented ssRNA genome ~28-32 Kb ~14 Kb, 10-14 proteins

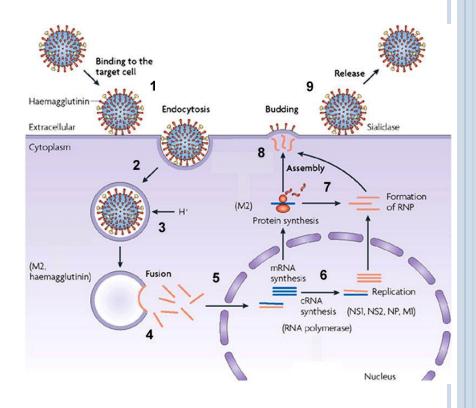
Coronavirus and influenza virus replication cycles

Coronavirus

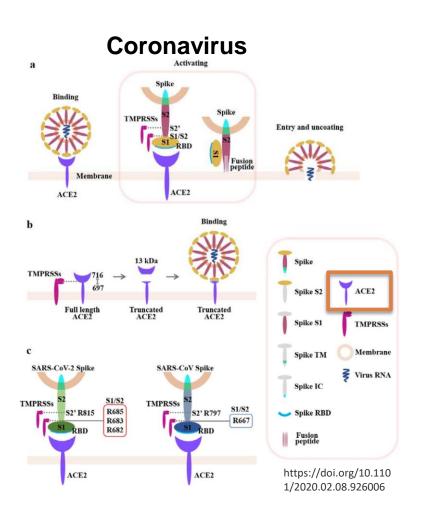
5. Virion Assembly

ER-Golgi Intermediate Compartment

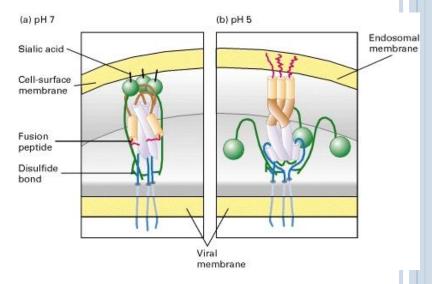
Influenza virus



DISTINCT RECEPTOR BINDING FEATURES OF SARS VS. INFLUENZA VIRUSES

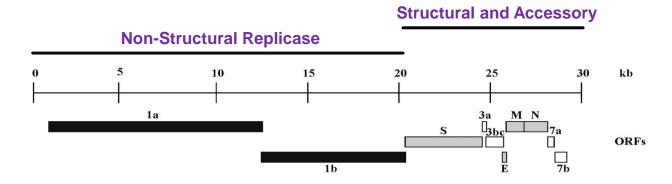


Influenza virus



Influenza HA binds to sialic acid residues on diverse surface proteins

Coronavirus Genome Encodes Several IFN Antagonists



1. Non-Structural Proteins (nsp1-16)

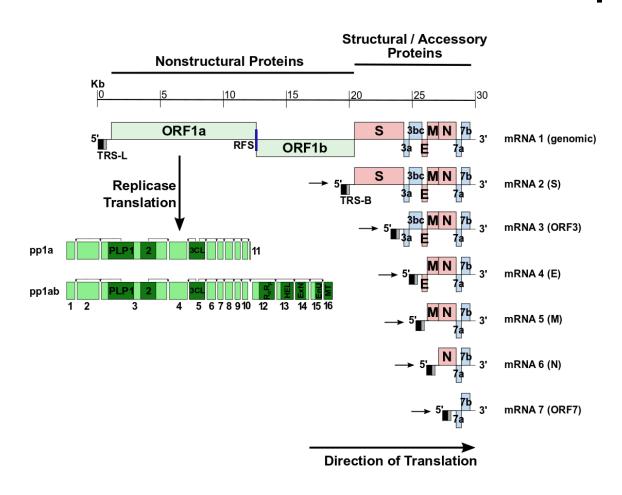
Conserved across CoVs Various, required functions IFN antagonists: nsp1, PLP2

(nsp3)

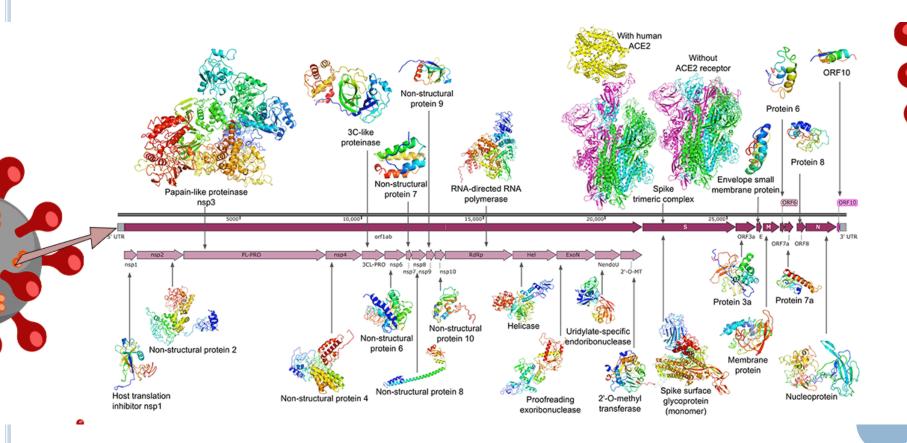
2. Accessory Proteins

Unique to subfamilies and species Function dispensable for replication Encode virulence factors

Coronavirus Genome Structure and Duplication



LARGE SARS-COV-2 PROTEOME CONTAINS MANY IMMUNOMODULATORY NON-STRUCTURAL PROTEINS



SARS-CoV-2 VS. INFLUENZA VIRUS SUMMARY

SARS-CoV-2

- RNA virus (+ sense)
- Single segment
- Large genome
- Multiple immune antagonists
- Specific receptor (ACE2)

Influenza virus

- RNA virus (- sense)
- 8 segments
- Much smaller genome (than CoV)
- Single immune antagonist (ds RNA sequestration)
- Non-specific receptor